

**LEARNING UNLIMITED AS BIOLOGY PRACTICALS WEEKLY SCHEDULE**

WEEK/PRACTICAL	SYLLABUS OBJECTIVE	PRACTICAL ACTIVITIES
<p>1. BIOLOGICAL MOLECULES (IDENTIFICATION)</p>	<p>a. carry out tests for reducing sugars and non-reducing sugars, the iodine test for starch, the emulsion test for lipids and the biuret test for proteins to identify the contents of solutions</p>	<p>You are required to carry out tests, using only the reagents provided, to identify each of the solutions S1, S2 and S3.</p> <p>One of the solutions is glucose, another a protein and the third a carbohydrate other than glucose.</p> <p>You are required to identify each of the solutions, S1, S2 and S3. You must use only the reagents provided.</p> <p>9700/31/M/J/08</p>
<p>2. BIOLOGICAL MOLECULES (QUALITATIVE)</p>	<p>a. carry out a semi-quantitative Benedict's test on a reducing sugar using <b>SIMPLE</b> dilution, standardising the test and using the results (colour standards or time to first colour change) to estimate the concentration</p>	<p>You will need to:</p> <ul style="list-style-type: none"> <li>• prepare known concentrations of sucrose solution using simple (proportional) dilution</li> <li>• carry out the non-reducing sugar test on these concentrations</li> <li>• carry out the non-reducing sugar test on the unknown concentration of sucrose solution in fruit extract, U</li> <li>• estimate the concentration of sucrose in fruit extract, U.</li> </ul> <p>9700/33/M/J/18</p>
<p>3. BIOLOGICAL MOLECULES (QUANTITATIVE)</p>	<p>a. carry out a semi-quantitative Benedict's test on a reducing sugar using <b>SERIAL</b> dilution, standardising the test and using the results (colour standards or time to first colour change) to estimate the concentration</p>	<p>You are required to:</p> <ul style="list-style-type: none"> <li>• prepare different concentrations of the glucose solution G</li> <li>• carry out the Benedict's test on each of the concentrations of glucose solution you have prepared</li> <li>• carry out the Benedict's test on the solution representing blood plasma P</li> <li>• use the results of the Benedict's tests to estimate the concentration of glucose in P.</li> </ul> <p>9700/35/M/J/16</p>

<p>4. ENZYMES 1 (INHIBITORY/CONCENTRATION)</p>	<p>a. investigate the progress of an enzyme-catalysed reaction by measuring rates of formation of products (for example, using catalase) or rates of disappearance of substrate (for example, using amylase)</p> <p>b. investigate and explain the effects of the following factors on the rate of enzyme-catalysed reactions:</p> <ul style="list-style-type: none"> <li>• enzyme concentration</li> <li>• substrate concentration</li> <li>• inhibitor concentration</li> </ul>	<p>1. You are required to investigate the effect of different concentrations of ethanol on the activity of yeast cells by measuring the change in pH, using Universal Indicator paper.</p> <p>2. Simple Dilution</p> <p>9700/31/M/J/09</p>
<p>5. ENZYMES 2 (pH/ TEMPERATURE)</p>	<p>a. investigate and explain the effects of the following factors on the rate of enzyme-catalysed reactions:</p> <ul style="list-style-type: none"> <li>• temperature</li> <li>• pH (using buffer solutions)</li> </ul>	<p>You are required to investigate the effect of temperature (independent variable) on the activity of a yeast cell suspension.</p> <p>9700/32/M/J/16</p>
<p>6. FULL MICROSCOPY SESSION</p>	<p>a. compare the structure of typical animal and plant cells by making temporary preparations of living material and using photomicrographs</p> <p>b. calculate the linear magnifications of drawings, photomicrographs and electron micrographs</p> <p>c. use an eyepiece graticule and stage micrometer scale to measure cells and be familiar with units (millimetre, micrometre, nanometre) used in cell studies</p> <p>d. calculate actual sizes of specimens from drawings, photomicrographs and electron micrographs observe and draw the mitotic stages visible in temporary root</p> <p>e. tip squash preparations and in prepared slides of root tips of species such as those of <i>Vicia faba</i> and <i>Allium cepa</i></p>	<ul style="list-style-type: none"> <li>• SIZE</li> <li>• SCALE</li> <li>• DRAWINGS</li> <li>• CALIBRATION</li> <li>• CALCULATIONS</li> </ul> <p>CONTENT EXTRACTED FROM SEVERAL PAST PAPERS</p>
<p>7. OSMOSIS</p>	<p>a. investigate diffusion and osmosis using plant tissue and non-living materials, such as Visking tubing and agar</p> <p>b. investigate the effects of immersing plant tissues in solutions of different water potentials, using the results to estimate the water potential of the tissues</p>	<p>1. You are required to investigate how much glucose diffuses through selectively permeable Visking (dialysis) tubing in 15 minutes.</p> <p>2. 9700/33/M/J/10</p>

<p>8. LIVE SPECIMEN</p>	<p>a. Draw the distribution of different <b>LIVE</b> tissues in plant and animal specimens and label the drawings appropriately</p>	<p>1. You are required to:</p> <ul style="list-style-type: none"> <li>• observe and record the cells in S1 and the effect of adding water, W, to these cells</li> <li>• observe and record the onion cells to identify the concentration of the sodium chloride solutions, S2 and S3.</li> </ul> <p>2. You are required to investigate the effect of staining the tissues of sweet banana with iodine solution.</p> <p>9700/31/M/J/12</p>
<p>9. TRANSPIRATION</p>	<p>a. transpiration rate using simple potometers, leaf impressions, epidermal peels and grids for determining surface area</p> <p>b. make annotated drawings, using prepared slides of cross-sections, to show how leaves of xerophytic plants are adapted to reduce water loss by transpiration</p> <p>c. draw and label from prepared slides plan diagrams of transverse sections of stems, roots and leaves of herbaceous dicotyledonous plants using an eyepiece graticule to show tissues in correct proportions</p> <p>d. draw and label from prepared slides the cells in the different tissues in roots, stems and leaves of herbaceous dicotyledonous plants using transverse and longitudinal sections</p> <p>e. draw and label from prepared slides the structure of xylem vessel elements, phloem sieve tube elements and companion cells and be able to recognise these using the light microscope</p>	<p>You are provided with three samples, <b>S1</b>, <b>S2</b> and <b>S3</b> from plant root extracts, removed at different times of the year:</p> <ul style="list-style-type: none"> <li>• the middle of winter, when there was no plant growth</li> <li>• the beginning of spring, when plant growth begins</li> <li>• the middle of spring, during active growth.</li> </ul> <p>9700/35/M/J/14</p>
<p>10. IMMOBILISATION OF BIOLOGICAL MOLS/ENZYMES</p>	<p>a. investigate and explain the effect of immobilising an enzyme in alginate on its activity as compared with its activity when free in solution</p>	<p>You are required</p> <ul style="list-style-type: none"> <li>• to immobilise the yeast cells in sodium alginate beads</li> <li>• to follow a student's procedure to investigate the independent variable, changing the surface area of the beads.</li> </ul> <p>9700/32/M/J/10</p>

<p>11. MOVEMENT OF SUBSTANCES ACROSS MEMBRANE</p>	<p>c. calculate surface areas and volumes of simple shapes (including cubes) to illustrate the principle that surface area to volume ratios decrease with increasing size</p> <p>d. investigate the effect of changing surface area to volume ratio on diffusion using agar blocks of different sizes</p>	<p>You are required to investigate the effect of concentration of hydrochloric acid (independent variable) on the rate of diffusion.</p> <p>The investigation involves placing agar cubes containing an indicator into dilute hydrochloric acid.</p> <p>As the acid diffuses into the agar cubes the indicator changes colour.</p> <p>9700/32/M/J/18</p>
<p>12. TITRATION</p>	<p>a. Estimate the concentration of unknown solutions from qualitative results</p>	<p>Manufacturers of fruit juice need to know the concentration of ascorbic acid (vitamin C) in fruit.</p> <p>The fruit is picked when the juice contains the highest concentration of ascorbic acid.</p> <p>The concentration of ascorbic acid can be estimated in a fruit extract by carrying out a test using starch solution and iodine solution.</p> <p>Iodine solution will be added one drop at a time using a syringe.</p> <p>9700/34/O/N/18</p>